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Corni Fructus: a review of chemical constituents and pharmacological activities

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Abstract

Cornus officinalis Sieb. et Zucc. is part of the genus Cornus of the family Cornaceae. Ripening and dry fruits (Corni Fructus) are recognized as an essential herb medicine in the traditional Chinese medicine (TCM) and have been widely used for over 2000 years. This review provides a comprehensive summary of Corni Fructus (CF), including the botany, phytochemistry, traditional use, and current pharmacological activities. According to the basic theory of TCM, CF usually participates in various Chinese medicinal formulae to exert the essential roles in replenishing liver and kidney, arresting seminal emission and sweat. Based on modern pharmacological studies, about 90 compounds have been isolated and identified from CF. In vivo and in vitro experimental studies indicate that CF exhibits extensive pharmacological activities including hypoglycemic, antioxidant, anti-inflammatory, anticancer, neuroprotective, hepatoprotective, and nephroprotective activities. However, only about 18% of chemical constituents in CF were tested. It means the potential pharmacological activities and clinical values of CF need to be further investigated.

Keywords: Cornus officinalis Sieb. et Zucc., Corni Fructus, Shan Zhu Yu, Phytochemistry, Pharmacological activity

Background

Cornus officinalis Sieb. et Zucc., commonly known as Shan Zhu Yu/山茱萸 (in Chinese), Asiatic Dogwood, and Japanese Cornel Dogwood, is a deciduous shrub or dungarunga in the genus *Cornus* (family *Cornaceae*). It is a heliophilous plant that grows in the warm-temperate zone. The most suitable growth temperature is between 20 and 30 °C, it also has a specific cold resistance that can temporarily grow in -18 °C low-temperature zone. *Cornus officinalis* Sieb. et Zucc. can be found in Anhui, Gansu, Jiangsu, Jiangxi, Shandong, Shanxi in China, Korea, and Japan. It usually grows in 400–1500 m high mountain slope, forest or forest edge. Ripening fruits are picked during September and October and dried in the air for medical uses [1, 2].

About 2200 years ago, Cornus officinalis Sieb. et Zucc. fructus (usually known as Corni Fructus) was first recorded in Shen Nong's Materia Medica (Fig. 1).

According to the basic theory of TCM, CF is characterized as replenishing liver and kidney, arresting seminal emission and sweat for its sour, astringent, and tepid properties [1]. It is used to treat four series of clinical symptoms. The first part of symptoms contains vertigo, tinnitus, weakness of the waist and knees which are caused by liver and kidney deficiency. CF is usually combined with Radix Rehmanniae Praeparata, Dioscoreae Rhizoma, Alismatis Rhizoma, Moutan Cortex, Poria to make Liuwei Dihuang Wan (六味地黄丸) replenish liver and kidney Yin [3]. For patients with kidney Yang deficiency, CF helps Cinnamomi Cortex, Aconiti Lateralis Radix Praeparata to reinforce Yang from Yin, e.g., Jingui Shenqi Wan (金匮肾气丸) [4]. The second part of symptoms contains spermatorrhoea and polydipsia. For patients with kidney deficiency, CF is frequently used with Radix Rehmanniae Praeparata, Dioscoreae Rhizoma, Cervi Cornu Pantotrichum, Psoraleae Fructus. For patients with dysfunction of the urinary bladder, CF is often applied with Mantidis Oötheca, Rubi Fructus, Rosae Laevigatae Fructus. The third part of symptom contains hypermenorrhea. CF is usually combined with Radix Rehmanniae Praeparata, Angelica Sinensis, Radix Paeoniae Alba to make Guchong Tang (固冲汤) preserve



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Fig. 1 Corni Fructus: a crude fruits, b processed fruits

Primordial Qi and stop Blood [5]. The fourth part of symptoms contains profuse cold sweating, pale complexion, cold limbs, and a feeble pulse. For patients with the Yang depletion syndrome, *Ginseng Radix et Rhizoma, Aconiti Lateralis Radix Praeparata*, and CF are applied in Laifu Tang (来复汤) to restore Yang from collapse. Medical practices indicate that CF can be combined with either Yin-tonifying or Yang-invigorating herbs to act as the sovereign drug or adjuvant drug in Chinese medicinal formulae and treat different types of TCM syndromes. Besides, CF is primarily made into the honey bolus to treat chronic diseases while is usually made into the decoction to treat acute conditions.

Chemical constituents

About 90 compounds have been isolated and identified from CF, including terpenoids, flavonoids, tannins, polysaccharides, phenylpropanoids, sterols, carboxylic acids, furans, and mineral substances. Chemical constituents are listed in Table 1. Among them, iridoids, tannins, and flavonoids are the major components. Their chemical structures are shown in Figs. 2, 3, 4 and 5.

Terpenoids (1-26) and flavonoids (27-39)

Most terpenoids and flavonoids in CF shared two similar isolation processes. Firstly, CF was percolated with ethanol to acquire the solvent which was then evaporated under reduced pressure. The resulting extract was suspended in water and then partitioned with ethyl acetate for several times. Finally, the extract was subjected to column chromatography over silica gel to yield compounds. Secondly, CF was grounded into powder and then subjected to supercritical carbon dioxide to yield extract. The resulting extract was subjected to GC–MS to identify the chemical components. So far, 26 terpenoids and 13 flavonoids have been isolated and identified from

CF. Among terpenoids, the pharmacological activities of sweroside (1), loganin (5), cornuside (23), ursolic acid (24), and oleanolic acid (25) have been further assayed, and a wide range of pharmacological activities has been revealed. Furthermore, two types of flavonoids namely kaempferol (28), quercetin (33), and their derivatives are the essential flavonoids.

Tannins (40-69)

During the isolation process, CF was firstly homogenized in acetone and then filtered to acquire an aqueous solution which was sequentially extracted with diethyl ether and ethyl acetate. The extract was subjected to column chromatography to give compounds. Finally, the chemical structure and molecular weight were determined using nuclear magnetic resonance (NMR) spectroscopy. To date, 30 tannic acids have been isolated from CF. Tsutomu HATANO identified 28 of them. Many tannic acids in this Chinese herb have the large molecular weight, e.g., the molecular weight of Cornusiins A–F and Camptothins A–B are even larger than 1000 Da [6, 7], because dimers and trimers exist in these types of tannic acids.

Polysaccharides (70-79)

Wu and Yin identified most polysaccharides in CF [8, 9]. In their isolation process, hot water or petroleum ether was initially used for combining with assistant ultrasonic and microwave to break the cell wall to isolate polysaccharides. Further separation and purification were achieved by the combination of several techniques, e.g., fractional precipitation, ethanol precipitation, ion-exchange chromatography and affinity chromatography. Finally, infrared spectroscopy analysis and morphological analysis were used to determine the physiochemical and structural features of the polysaccharide.

Table 1 Chemical constituents identified from CF

No.	Chemical class	Compound name	Chemical formula	Exact mass	References
	Terpenoids				
1	Iridoids	Sweroside	$C_{16}H_{22}O_{9}$	358.1264	[73]
2		Loganic acid	$C_{16}H_{24}O_{10}$	376.1369	[19]
3		Cornin	C ₁₇ H ₂₄ O ₁₀	388.1369	[73]
4		7-Dehydrologanin	$C_{17}H_{24}O_{10}$	388.1369	[74]
5		Loganin	$C_{17}H_{26}O_{10}$	390.1526	[74]
6		7-α-Morroniside	$C_{17}H_{26}O_{11}$	406.1475	[75]
7		7-β-Morroniside	$C_{17}H_{26}O_{11}$	406.1475	[75]
8		7-α-O-Methyl-morroniside	$C_{18}H_{28}O_{11}$	420.1632	[75]
9		7-β-O-Methyl-morroniside	$C_{18}H_{28}O_{11}$	420.1632	[75]
10		7-α-O-Ethyl-morroniside	$C_{18}H_{29}O_{11}$	421.1710	[75]
11		7-β- <i>O</i> -Ethyl-morroniside	$C_{18}H_{29}O_{11}$	421.1710	[75]
12		7-α- <i>O</i> -Butyl-morroniside	$C_{21}H_{34}O_{11}$	462.2101	[43]
13		7-β-O-Dimethyl-butanedioate morroniside	$C_{23}H_{34}O_{15}$	550.1898	[76]
14		Logmalicids A	$C_{21}H_{29}O_{14}$	505.1557	[24]
15		Logmalicids B	$C_{21}H_{29}O_{14}$	505.1557	[24]
16		Cornusfuroside A	$C_{25}H_{34}O_{13}$	542.1999	[77]
17		Cornusfuroside B	$C_{25}H_{34}O_{13}$	542.1999	[77]
18		Cornusfuroside C	$C_{25}H_{34}O_{13}$	542.1999	[77]
19		Cornusfuroside D	$C_{29}H_{34}O_{15}$	622.1898	[77]
20	Secoiridoids	Linalool	C ₁₀ H ₁₈ O	154.1358	[78]
21		Linalool oxide	$C_{10}H_{18}O_2$	170.1307	[79]
22		Secoxyloganin	$C_{17}H_{24}O_{11}$	404.1319	[24]
23		Cornuside	$C_{24}H_{30}O_{14}$	542.1636	[74]
24	Triterpenoids	Ursolic acid	$C_{30}H_{48}O_3$	456.3603	[74]
25		Oleanolic acid	$C_{30}H_{48}O_3$	456.3603	[15]
26		Arjunglucoside II	$C_{36}H_{58}O_{10}$	650.4030	[74]
	Flavonoids				
27		Naringenin	$C_{15}H_{12}O_5$	272.0685	[74]
28		Kaempferol	$C_{15}H_{10}O_6$	286.0477	[79]
29		Kaempferide	$C_{16}H_{12}O_6$	300.0634	[80]
30		Kaempferol-3- <i>O</i> -β-D-galactopyranoside	$C_{21}H_{20}O_{11}$	448.1006	[24]
31		Kaempferol-3- <i>O</i> -β-D-glucoside	$C_{21}H_{20}O_{11}$	448.1006	[74]
32		Kaempferol-3- <i>O</i> -β-D-rutinoside	$C_{27}H_{30}O_{15}$	594.1585	[24]
33		Quercetin	$C_{15}H_{10}O_{7}$	302.0427	[79]
34		Quercetin-3- <i>O</i> -β-D-galactopyranoside	$C_{21}H_{20}O_{12}$	464.0955	[24]
35		Quercetin-3-O-β-p-glucuronide	C ₂₁ H ₁₈ O ₁₃	478.0747	[24]
36		Quercetin-3-O-β-p-glucuronide methyl ester	$C_{22}H_{20}O_{13}$	492.0904	[24]
37		Quercetin-3-O-β-D-(6-n-butyl glucuronide)	$C_{25}H_{25}O_{13}$	533.1295	[15]
38		(—)-Epicatechin-3- <i>O</i> -gallate	$C_{22}H_{18}O_{10}$	442.0900	[15]
39		Isoquercitrin	$C_{21}H_{20}O_{12}$	464.0955	[80]

Table 1 (continued)

No.	Chemical class	Compound name	Chemical formula	Exact mass	References
	Tannins				
40		Gallic acid	$C_7H_6O_5$	170.0215	[74]
41		7-O-Galloyl-D-sedoheptulose	C ₁₄ H ₁₈ O ₁₁	362.0849	[19]
42		Gemin D	$C_{27}H_{22}O_{18}$	634.0806	[6, 7]
43		Oenothein C	$C_{34}H_{24}O_{22}$	784.0759	[6, 7]
44		3-O-Galloyl-p-glucose	$C_{13}H_{16}O_{10}$	332.0743	[6, 7]
45		2,3-Di- <i>O</i> -galloyl-p-glucose	$C_{20}H_{20}O_{14}$	484.0853	[6, 7]
46		1,2,3-Tri- <i>O</i> -galloyl-β-p-glucose	$C_{27}H_{24}O_{18}$	636.0963	[6, 7]
47		1,2,6-Tri- <i>O</i> -galloyl-β-p-glucose	$C_{27}H_{24}O_{18}$	636.0963	[6, 7]
48		1,2,3,6-Tetra-O-galloyl-β-p-glucose	$C_{34}H_{28}O_{22}$	788.1072	[6, 7]
49		Tellimagrandin I	$C_{34}H_{26}O_{22}$	786.0916	[6, 7]
50		Tellimagrandin II	C ₄₁ H ₃₀ O ₂₆	938.1025	[6, 7]
51		Isocoriariin F	C ₃₄ H ₂₆ O ₂₃	802.0865	[6, 7]
52		Coriariin F	C ₃₄ H ₂₆ O ₂₃	802.0865	[6, 7]
53		Rugosin B	C ₄₁ H ₃₀ O ₂₇	954.0974	[6, 7]
54		Isorugosin B	C ₄₁ H ₃₀ O ₂₇	954.0974	[6, 7]
55		Isoterchebin	C ₄₁ H ₃₀ O ₂₇	954.0974	[6, 7]
56		Isorugosin A	C ₄₈ H ₃₄ O ₃₁	1106.1084	[6, 7]
57		Rugosin D	C ₈₂ H ₅₈ O ₅₂	1874.1894	[6, 7]
58		Isorugosin D	C ₈₂ H ₅₈ O ₅₂	1874.1894	[6, 7]
59		Camptothin A	C ₆₁ H ₄₆ O ₄₀	1418.1565	[6, 7]
60		Camptothin B	C ₇₅ H ₅₄ O ₄₈	1722.1785	[6, 7]
61		Cornusiin B	C ₄₈ H ₃₀ O ₃₀	1086.0822	[6, 7]
62		Cornusiin A	C ₆₈ H ₅₀ O ₄₄	1570.1675	[6, 7]
63		Cornusiin D	C ₇₅ H ₅₄ O ₄₈	1722.1785	[6, 7]
64		Cornusiin E	C ₈₂ H ₅₈ O ₅₂	1874.1894	[6, 7]
65		Cornusiin F	C ₉₅ H ₇₀ O ₆₂	2202.2325	[6, 7]
66		Cornusiin C	C ₁₀₂ H ₇₄ O ₆₆	2354.2434	[6, 7]
67		Methyl tri-O-methylgallate	C ₁₁ H ₁₄ O ₅	226.0841	[6, 7]
68		Dimethyl hexamethoxydiphenate	$C_{22}H_{26}O_{10}$	450.1526	[6, 7]
69		Trimethyl-octa- <i>O</i> -methylvaloneate	C ₃₂ H ₃₆ O ₁₅	660.2054	[6, 7]
0,5	Polysaccharides	eury. deta d'interry.valdireate	-32' '36 - 15	000.203	[0].]
70	,	Co-4			[8]
71		COP-1			[9]
72		COP-2			[9]
73		COP-3			[9]
74		COP-4			[9]
75		FCAP1			[81]
76		FCP5-A			[8]
77		PFCA-III			[8]
78		PFCC-I			[8]
79		SZYP-2			[8]
, ,	Other compounds	JEII Z			ί∩Ì
80	Phenylpropanoids	<i>p</i> -Hydroxycinnamic acid	СНО	164.0473	[74]
	i rieriyipiopariolus	Caffeic acid	C ₉ H ₈ O ₃	180.0423	
81		Сапеіс асіd Caftaric acid monomethyl ester	$C_9H_8O_4$		[15]
82			C ₁₄ H ₁₄ O ₉	326.2556	[15]
83	Sterols	Caffeoyltartaric acid dimethyl ester β-Sitosterol	C ₁₅ H ₁₆ O ₉	340.0794	[76]
84	SIGIOIZ		C ₂₉ H ₅₀ O	414.7067	[15]
85		Daucosterol-6'-malate	C ₃₉ H ₆₄ O ₁₀	692.4499	[80]

Table 1 (continued)

No.	Chemical class	Compound name	Chemical formula	Exact mass	References
86	Carboxylic acids	Succinic acid	C ₄ H ₆ O ₄	118.0266	[85]
87		Malic acid	$C_4H_6O_5$	134.0215	[85]
88		Methylmalic acid	$C_5H_8O_5$	148.0372	[74]
89		Citric acid	$C_6H_8O_7$	192.0270	[85]
90		Butoxysuccinic acid	$C_8H_{14}O_5$	190.1938	[15]
91	Furans	5-Hydroxymethylfurfural	$C_6H_6O_3$	126.0317	[73]
92		Dimethyltetrahydrofuran cis-2,5-dicarboxylate	C ₈ H ₁₂ O ₅	188.0685	[79]
93	Mineral substances	Ca, Fe, K, Mg, Mn, Zn			[82]

Other compounds (80-93)

Four phenylpropanoids, two sterols, five carboxylic acids, two furans, and several mineral substances have also been determined. Among them, 5-hydroxymethylfurfural exhibits diverse biological activities. Besides, Chen, Li, and Wen identified 32, 16, and 48 volatile compounds by GC–MS, respectively [10–12].

Pharmacological activities

Although just a few chemical constituents from CF are assayed for their biological activities, these components displayed diverse pharmacological activities. Detailed biological activities are summarized in Table 2.

Hypoglycemic activity and diabetic target organs protective activity

Diabetes mellitus (DM) is a group of long-term and chronic metabolic disorders which are associated with high serum glucose levels. Compared with the no treatment diabetic animal model group, CF extract (at 300 mg kg^{-1} 2 day^{-1} and 400 mg kg^{-1} day^{-1} p.o.), loganin, morroniside, and ursolic acid (each at 200 mg kg⁻¹ day⁻¹ p.o.) for 4 weeks can significantly decrease fasting blood glucose and alleviate polyphagia, polydipsia, polyuria, and weight loss [13, 14]. In He's study, metformin (at 200 mg kg⁻¹ day⁻¹ p.o.) demonstrated better effect [14]. Besides, loganin, morroniside, ursolic acid, and butyl morroniside (each at 100 μ mol L⁻¹) can protect the pancreatic β -cells from high glucose-induced excessive oxidative stress and apoptosis [14, 15], may further increase the insulin release. Compared with the insulin treatment, CF extract, (-)-epicatechin-3-O-gallate, and caftaric acid monomethyl ester (each at 50 µmol L⁻¹) can also significantly inhibit α-glucosidase activity to slow down the elevation of serum glucose levels [14, 16, 17] and suppress the hepatic gluconeogenesis by decreasing the protein and mRNA levels of PEPCK in vitro [15, 18].

Also, CF extract, iridoid glycosides, and the single compound can decrease 24 h urine protein and serum levels of urea nitrogen and creatinine. To be specific, loganin, morroniside, and 7-*O*-galloyl-D-sedoheptulose (each at 20–100 mg kg⁻¹ day⁻¹ p.o.) for 10 days and 8 weeks can significantly inhibit both AGE/RAGE formation [19–22] and CTGF production [23] in db/db mice or STZ-induced diabetic nephropathy model. They can also significantly alleviate diabetic organ injury by decreasing the production of NF-κB and its downstream synthetases and cytokines [19–25], increasing antioxidant enzyme production [19, 26, 27], and suppressing apoptotic cell death [27, 28].

Antioxidant activity

Long-term oxidative stress will generate excessive ROS to oxidize protein, lipids, DNA and then cause cell death, tissue damage, and organ dysfunction. Ideal antioxidant drugs are required to regulate the defense system and scavenge excessive ROS. Studies indicated that morroniside (at 1, 10, 100 μ mol L⁻¹) for 24 h and total saponins (at 60 and 120 mg kg⁻¹ day⁻¹ p.o.) for 4 weeks regulated Ca²⁺ and NO release [29, 30], the aqueous extract (at 0.25-2.0 mg mL⁻¹) for 20 h modulated GSH redox cycle [31], the aqueous extract, the ethanol extract (at 0.01-0.1 mg mL⁻¹), morroniside (at $0.05-2 \mu g \text{ mL}^{-1}$), and ursolic acid (at $0.05-2 \mu g \text{ mL}^{-1}$) for 24 h promoted antioxidant enzymes syntheses [31– 33] to inhibit lipid peroxidation [29], 5-hydroxymethylfurfural (at 100-400 µmol L⁻¹) for 3 days decreased ROS release [34], morroniside (at 100 μ mol L⁻¹) for 2 days recovered cell cycle to normal state [35]. Mentioned effects significantly together reduced the oxidative stress-induced damages compared with the no treatment group.

Table 2 Summary of pharmacological activities of CF

Extracts or compounds	Disease models	Specific effects	References
Hypoglycemic activity			
Oleanolic acid	Fasting rat	Decrease plasma glucose levels. Regulate ACh release from nerve terminals to activate muscarinic M3 receptors in the pancreatic cells and increase C-peptide and insulin release	[83]
Iridoid glycosides	STZ-induced rat as DM model	Show a-glucosidase inhibition activity in vitro and decrease serum glucose levels in vivo	[16]
Loganin Morroniside Ursolic acid	STZ-induced mice as DM model, HepG2 cell lines	Show a-glucosidase inhibition activity in vitro. Decrease fasting blood glucose and alleviate weight loss, polydipsia, and polyphagia. Increase SOD activity and ROS scavenging activity. Attenuate aldose reductase activity and decrease MDA plasma level and renal somatic indices in mice	[14]
Butyl morroniside (—)-Epicatechin-3-0-gallate Caftaric acid monomethyl ester	High glucose-induced BRIN-BD11 and H4IIE cell lines as in vitro DM model	Increase glucose uptake efficiency. Reduce PEPCK mRNA level and NO production. Inhibit pancreatic β -cell death	[15]
Aqueous extract	STZ-induced rat as diabetic organs injury model	Decrease levels of glucose and TC in serum, and α-SMA expression in kidney. Improve the pathohistological injury of pancreas, kidney, lung, and liver	[13]
Aqueous extract	Normal rat	Show a-glucosidase inhibition activity in vitro, and exhibit hypoglycemic effect via oral sucrose tolerance test in vivo	[17]
Aqueous extract	Dexamethasone and 8-bromo-cAMP-induced BRIN-BD11 and H4IIE cell lines as in vitro DM model	Increase insulin release. Decrease PEPCK mRNA level	[18]
Nephroprotective activity			
Loganin	STZ-induced rat and high glucose-induced HK-2 as in vivo and in vitro diabetic nephropathy model	Improve renal function. Decrease CTGF level in kidney and serum via ERK signaling pathway	[23]
Morroniside Loganin 7-O-Galloyl-p-sedoheptulose	Db/db mice as obesity-associated type 2 diabetic nephropathy model	Suppress formation of AGEs and TBARS in the kidney, Reduce the production of SREBP-182, NF-kB p65, COX-2, and iNOS. Decrease GSH/GSSG ratio and levels of serum glucose, TC, and TG	[19]
7-O-Galloy/-p-sedoheptulose	STZ-induced rat as diabetic nephropathy model	Decrease serum creatinine, renal glucose, and urinary protein. Reduce the production of AGE, RAGE, HO-1, intracellular glycation, CML, GA- pyridine, and TBARS	[21]
Iridoid glycosides	STZ-induced rat as diabetic nephropathy model	Suppress over-deposition of fibronectin and laminin in the kidney. Reduce protein and mRNA levels of TGF- β 1 in serum and glomeruli	[25]
Iridoid glycosides Triterpene acids	Db/db mice as obesity-associated type 2 diabetic nephropathy model	Improve the histological injury of kidney and pancreas. Ameliorate the structural alterations in mesangial cells and the podocytes in the renal cortex. Inhibit ECIM accumulation in the kidney. Decrease 24 h urine protein and serum levels of urea nitrogen and creatinine. Increase insulin release, and decrease fasting blood glucose and levels of TC, TG, and GSP. Attenuate food consumption, water intake, and urine volume. Reduce the production of RAGE, NF-κΒ, SphK1, and TGF-β	[22]
CF extract	STZ-induced rat as diabetic nephropathy model	Inhibit AGE formation in the kidney. Attenuate hyperglycemia and proteinuria. Reduce the production of RAGE, NF- κ B, TGF- β 1, and CML	[20]
			l

Table 2 (continued)			
Extracts or compounds	Disease models	Specific effects	References
Ethanol extract	High glucose-induced mesangial cells as in vitro diabetic nephropathy model	Decrease the production of Col V, FN, and IL-6	[24]
Myocardial protection activity			
Morroniside	High glucose-induced rat as diabetic cardiomyopathy model	Inhibit myocardial cell apoptosis. Elevate Bd-2 production and decrease expressions of Bax and caspase-3	[28]
Triterpene acids	STZ-induced rat as diabetic cardiomyopathy model	Inhibit the ventricular remodeling and regulate the systolic and diastolic function of the left ventricle. Increase insulin release and reduce serum glucose levels. Enhance GSX and SOD activity. Increase the production of calstabin 2, PLB, and SERCA2a. Decrease protein and mRNA levels of ECE, iNOS, MDA, ET-1, and propreET-1	[26]
Testis-protective activity			
Iridoid glycosides	STZ-induced rat as diabetic testicular damage model	Improve the pathohistological injury of testes and pancreas. Increase serum insulin release and decrease blood glucose levels. Alleviate weight loss, polydipsia, polyphagia, and polyuria. Increase CAT and SOD activity. Reduce the production of AGEs, RAGE, ROS, MDA, and p-p38 MAPK. Down-regulate Bax/Bcl-2 ratio and spermatogenic cell apoptosis	[27]
Antioxidant activity			
Morroniside	Hydrogen peroxide-induced SH-SYSY cell line as in vitro neurodegenerative disorder model	Suppress intracellular accumulation of Ca ²⁺ . Increase SOD activity and reduce the loss of MMP. Inhibit cytotoxicity	[59]
Morroniside	High ambient glucose-induced endothelial cell injury model	Attenuate cellular morphological damage. Repair cell cycle progression and improve cell viability	[35]
Ursolic acid	Hydrogen peroxide-induced HEI-OC1 cell line as in vitro inner ear diseases model	Increase antioxidant enzymes expressions, e.g., CAT and GPX. Suppress lipid peroxidation	[32]
5-Hydroxymethylfurfural	High glucose-induced HUVECs as in vitro oxidative stress model	Decrease levels of ROS, IL-8, JNK1, and JNK2/3. Increase P-Akt production	[34]
Total saponins	STZ-induced rat as a diabetic oxidative stress model	Regulate NO release and endothelium-dependent relaxation on the mesenteric artery. Reduce blood glucose levels	[30]
Aqueous extract	Hypoxanthine and xanthine oxidase-induced bovine PAECs as in vitro oxidative stress model	Regulate GSH redox cycle. Increase the intracellular GSH production and the activity of GSH peroxidase and GSH disulfide reductase. Reduce the intracellular level of GSH disulfide. Increase CAT and SOD activity and inhibit the production of hydrogen peroxide and superoxide anion	[31]
Ethanol extract	LPS-induced RAW 264.7 macrophage cells as in vitro oxidative stress model	Attenuate xanthine oxidase activity and ROS production. Induce the production of antioxidant enzymes, e.g., CAT, GSX, Cu/Zn-SOD, and Mn-SOD	[33]
Anti-inflammatory activity			
Cornuside	TNF- α -induced HUVECs as in vitro inflammation model	Decrease the production of ICAM-1, VCAM-1, MCP-1, and NF-xB. Inhibit NF-xB p65 translocation	[36]
Cornuside	LPS-induced RAW 264.7 macrophage cells as in vitro inflammation model	Decrease the production of COX-2, iNOS, PGE ₂ , NO, IL-1β, IL-6, and TNF-α. Suppress the translocation of NF-κB p65, the phosphorylation and degradation of IkB-α, and the phosphorylation of FRK1/2, JNK1/2, and p38	[38]

lable 2 (continued)			
Extracts or compounds	Disease models	Specific effects	References
Aqueous extract	LPS-induced RAW 264.7 macrophage cells as in vitro inflammation model	Decrease protein and mRNA levels of COX-2 and iNOS. Reduce PGE ₂ and NO production	[37]
Anticancer activity			
Aqueous extract	HSC-2, HSC-3, HSC-4, Ca9-22, NA cell lines as in vitro oral squamous cell carcinoma model	Produce broad radical peak under alkaline condition and increase the cytotoxicity and superoxide anion scavenging activity of vitamin C	[39]
Aqueous extract	E2-induced MCF-7 cell line as in vitro ER ⁺ human mammary carcinoma model	Inhibit cell line anchorage-independent growth and reduce the mitogenically inert metabolite E3 formation	[40]
Aqueous extract	Parental ER ⁺ MCF-7 cell line as in vitro human mammary carcinoma model	Suppress cell line anchorage-independent growth and induce G1 or G2/M arrest and apoptosis. Increase anti-proliferative E2 metabolites production	[41]
Aqueous extract	HepG2, SKHep1 and PLC/PRF/5 cell lines as in vitro hepatocellular carcinoma model	Inhibit cell proliferation. Exhibit free radicals scavenging activity and suppress lipid peroxidation and xanthine oxidase production	[42]
Neuroprotective activity			
Cornuside 1,2,3-Tri-O-galloyl-β-D-glucose 1,2,3,6-Tetra-O-galloyl-β-D-glucose Tellimagrandin I Tellimagrandin II	In vitro enzyme activities assay	Exhibit synergetic inhibitory activities against BACE1 and ChE	[45]
Morroniside	MCAO-induced rat as focal cerebral ischemia model	Decrease the infarction volume and improve neurological function. Increase GSH expression and SOD activity. Decrease the production and activity of MDA and caspase-3 in ischemic cortex tissues	[47]
5-Hydroxymethylfurfural	Hydrogen peroxide-induced rat hippocampal neurons as in vitro neurodegenerative disorder model	Enhance Bcl-2 production and suppress expressions of Bax, caspase-3, and p53	[84]
Iridoid glycosides	MCAO-induced rat as focal cerebral ischemia model	Improve neurological function. Increase the number of BrdU-positive cells and nestin-positive cells in the subventricular zone, and the number of new mature neurons and blood vessels in the striatum. Increase protein and mRNA levels of VEGF and FIK-1	[46]
Iridoid glycosides	Fimbria-fornix transected rat as cerebral ischemia model	Decrease neuron loss in the hippocampus and improve memory deficits. Increase the production of BDNF, NGF, BcI-2, SYP, Trk A, and GAP-43, and decrease the production of Bax and Cyt c	[49]
7R-O-Methyl-morroniside 7S-O-Methyl-morroniside 7-O-Butyl-morroniside Loganin Morroniside	Glutamate-induced HT22 cell lines as in vitro hippocampal cell injury	Improve cell viability	[43]

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Extracts or compounds	Disease models	Specific effects	References
Iridoid glycosides	Mycobacterium tuberculosis and guinea-pig myelin basic protein-induced experimental autoimmune encephalomyelitis rat as multiple sclerosis model	Increase the number of mature oligodendrocytes and reduce the number of oligodendrocyte progenitor cells. Inhibit the process of T cell entry to the central nervous system and attenuate microglia activation. Increase BDNF expression and decrease phosphorylation of JAK/ STAT1/3 and inflammatory cytokines production, e.g., IL-18, IFN-y, TNF-a	[48]
Iridoid glycosides	Mycobacterium tuberculosis and myelin oligodendrocyte glycoprotein- induced experimental autoimmune encephalomyelitis mouse as multiple sclerosis model	Decrease BDNF and NGF loss in the spinal cord	[50]
Aqueous extract	PC 12 cell lines	Increase cell neurite outgrowth. Inhibit extracellular Ca ²⁺ influx, and protein and mRNA levels of STIM1	[44]
Hepatoprotective activity 5-Hydroxymethylfurfural	Hydrogen peroxide-induced LO2 cell lines as in vitro hepatitis model	Promote S phase into G2/M phase and recover cell cycle to normal.	[51]
· · · · · · · · · · · · · · · · · · ·		Reduce NO production and caspase-4 activity and inhibit hepatocyte apoptosis	
5-Hydroxymethylfurfural	Hydrogen peroxide-induced L02 cell lines as in vitro hepatitis model	Improve hepatocyte morphology and reduce caspase-3&9 expressions	[52]
5-Hydroxymethylfurfural	D-Galactosamine/TNF-α-induced L02 cell lines as in vitro acute liver injury model	Inhibit hepatocyte apoptosis. Increase Bcl-2 production and decrease intracellular Ca ²⁺ level and production of ATF4, Bax, CHOP, PERK, and p-eIF2a	[53]
7-0-Galloyl-p-sedoheptulose	Db/db mice as obesity-associated type 2 diabetic liver injury model	Improve hepatic histological damage and decrease serum levels of ALT, AST, and blood glucose. Attenuate water intake, food consumption, and body weight gain. Decrease the production of AP-1, NF-xB p65, IL-6, TNF-q, ICAM-1, MCP-1, AGEs, RAGE, GA-pyridine, pentosidine, CEL, CMA, CML, leptin, resistin, p-ERK1/2, and p-JNK	[54]
Ethanol extract Improving osteoporosis activity	Acetaminophen-induced mice as liver injury model	Increase levels of CAT, HO-1, and SOD. Suppress lipid peroxidation	[55]
Sweroside	Rat osteoblasts and human MG-63 cell lines	Stimulate the osteocalcin secretion. Increase cell proliferation and inhibit apoptotic cell death. Increase ALP activity	[26]
CF extract	RANKL-induced mice BMDM as in vitro osteoclast differentiation model	Suppress osteoclast differentiation. Reduce protein and mRNA levels of c-Fos, NFATc1, OSCAR, and TRAP. Inhibit phosphorylation of p-38 and c-JNK and degradation of I+xB	[57]
Promoting melanogenesis activity			
Methanol extract	Melan-a cell lines	Increase the production and activity of tyrosinase. Increase MITF-M mRNA level and TRP-1&2 production	[58]
Immunomodulatory activity			
Aqueous extract	C57BL/6 mice are transplanted with a skin graft from Balb/C donors	Prolong skin allograft survival. Reduce the number of graft-infiltrating T cells and inhibit their proliferation. Decrease intracellular IL-12 expression by intragraft DCs and IFN-y expression by graft-infiltrating T cells. Reduce intragraft IL-12 mRNA level	[59]

Table 2 (continued)			
Extracts or compounds	Disease models	Specific effects	References
Lung-protective activity Oleanolic acid Ursolic acid	Epidermal growth factor—and phorbol ester-induced NCI-H292 cell lines Decrease protein and mRNA levels of MUC5AC mucin as in vitro airway diseases model	Decrease protein and mRNA levels of MUC5AC mucin	[60]
Aqueous extract	Ovalbumin-induced BALB/c mice as allergic asthma model	Inhibit eosinophil infiltration and ameliorate allergic airway inflammation [61] and airway hyperresponsiveness. Decrease the production of IL-5&13 and OVA-specific IgE	[61]
Vasorelaxation activity Cornuside	Phenylephrine-contracted rat aorta and HUVEC	Dilate vascular smooth muscle in the rat and increase cGMP production [62] in vitro	[62]
Antiviral activity Aqueous extract	CVA16 infected Vero cells as in vitro HFMD model	Inhibit CVA16 replication	[63]

Anti-inflammatory activity

Prolonged and incurable inflammation may cause many diseases, e.g., atherosclerosis, cancer, ulcerative colitis. In LPS and TNF- α -induced cell inflammation models, compared with the no treatment group, CF aqueous extract (at 0.2, 1, 5 mg mL⁻¹) and cornuside (at 1, 10, 50 μ mol L⁻¹) for 24 h significantly inhibited NF- κ B p65 translocation, down-regulated COX-2 and iNOS production, finally decreased PGE₂ and NO levels to control excessive inflammatory responses [36–38].

Anticancer activity

CF aqueous extract significantly enhanced both the cytotoxicity and superoxide anion scavenging activity of vitamin C at 0.5 and 36 μg mL⁻¹, respectively. Together with CF aqueous extract, vitamin C further inhibited proliferation and induced apoptosis in several human oral squamous cell carcinoma cell lines. Compared with no treatment, the proliferation inhibition rate was at 1.3–71.0% [39]. Furthermore, the aqueous extract (at 1.0 mg mL⁻¹) for 2 days significantly exhibited anti-ER⁺ human mammary carcinoma activity by inhibiting cell anchorage-independent growth, regulating the metabolism of E2 and E3 [40], and influencing cell cycle progression and cellular apoptosis [41]. Finally, the aqueous extract has been tested for its cancer inhibitory effect in several hepatocellular carcinomas and leukemic cell lines. The study indicated that the aqueous extract inhibited the tumor cell proliferation in a dose-dependent manner at 0.11-0.337 mg mL⁻¹, exhibited oxygen free radicals scavenging activity (at 50 μ g mL⁻¹), attenuated xanthine oxidase production (at 2.62 mg mL⁻¹) and lipid peroxidation (at 0.892 mg mL⁻¹) [42]. In this study, CF aqueous extract exhibited the similar effects compared with 5-fluorouracil (at 0.5, 1, 5 μ g mL⁻¹).

Neuroprotective activity

Many compounds in CF were further tested for the neuroprotective effects. 7R-O-Methyl-morroniside, 7S-O-methyl-morroniside, 7S-O-methyl-morroniside, 7S-O-methyl-morroniside, loganin, and morroniside (each at 10 and 50 $\mu\text{mol L}^{-1}$) for 1 h significantly protected the neurons against glutamate-induced neurotoxicity up to about 78% compared with the no treatment group [43]. CF aqueous extract (at $60~\mu\text{g mL}^{-1}$) significantly inhibited the extracellular Ca^{2+} influx to increase cell neurite outgrowth [44]. Also, cornuside, isoterchebin, and tellimagrandin II (each at $25\text{-}100~\mu\text{mol L}^{-1}$) displayed anti-Alzheimer's disease potential due to their synergetic inhibitory activities against BACE1 and ChE [45].

Cerebral ischemia, multiple sclerosis, and neurodegenerative disorder models are applied in

animal experiments. Iridoid glycosides (at 60 and 180 mg kg $^{-1}$ day $^{-1}$ p.o.) for 1–4 weeks and morroniside (at 90 and 270 mg kg $^{-1}$ day $^{-1}$ p.o.) for 3 days significantly decreased the infarction volume, increased the number of new mature neurons and blood vessels, and improved nervous system function [46, 47]. Also, iridoid glycosides (at 50–180 mg kg $^{-1}$ day $^{-1}$ p.o.) for 3–4 weeks can significantly promote NGF and BDNF production [48–50], and repair the abnormal functions of microglia, oligodendrocyte, and T cell to maintain the central nervous system homeostasis [48].

Hepatoprotective activity

In hepatitis cell models, 5-hydroxymethylfurfural (at 0.2–1 and 0.79 µmol L^{-1}) for 24 h has been shown to protect hepatocytes from H_2O_2 induced-cytotoxicity by significantly decreasing NO and intracellular Ca^{2+} levels, inhibiting abnormal production of apoptosis-related proteins and recovering back to regular cell cycle [51–53]. In hepatitis animal models, 7-O-galloyl-D-sedoheptulose (at 20 and 100 mg kg $^{-1}$ day $^{-1}$ p.o.) for 6 weeks and CF ethanol extract (at 100–500 mg kg $^{-1}$ day $^{-1}$ p.o.) for 1 week significantly decreased the serum marker enzymes of hepatic damage, weakened the oxidative stress by promoting antioxidant enzymes production and inhibiting lipid peroxidation, finally improved hepatic histological injury [54, 55].

Other pharmacological activities

In addition to the mentioned pharmacological activities, CF has also been reported to exert multiple bioactivities. Firstly, sweroside (at 7.5 µg mL⁻¹) for 1 week significantly promoted the proliferation and differentiation of osteoblasts via the regulation of osteocalcin [56]. Also, CF extract (at $0-100 \mu g mL^{-1}$) for 4 days significantly inhibited osteoclast differentiation in a dose-dependent manner via the inhibition of the signaling cascades NF-κB/c-Fos/NFATc1 to improve osteoporosis [57]. Secondly, CF methanol extract (at 3.125–12.5 µg mL⁻¹) for 3 days significantly up-regulated synthesis and activity of tyrosinase, raised TRP-1&2 translation associating with increasing transcription of MITF-M, finally promoted melanogenesis by 36.1% [58]. Thirdly, CF aqueous extract possesses immunomodulatory activity. In C57BL/6 mice that were transplanted with a skin graft from Balb/C donors, CF extract significantly prolonged skin allograft survival synergistically by suppressing Th1 response, promoting regulatory T cell generation, and enhancing its suppressive function [59]. Fourthly, CF shows lung-protective activity via two studies. In the cellular test, oleanolic acid (at 10 and 100 μmol L⁻¹) and ursolic acid (at 100 μ mol L⁻¹) for 30 min' pretreatment significantly down-regulated MUC5AC mucin whose excessive

level would impair airway defenses to cause serious airway diseases [60]. In an animal experiment, CF aqueous extract (at 50 and 200 mg kg $^{-1}$ 3 day $^{-1}$ p.o.) for 5 weeks significantly decreased the production of inflammatory mediators and reduced eosinophil infiltration, finally attenuated allergic airway inflammation and airway hyperresponsiveness [61]. Fifthly, cornuside significantly dilated vascular smooth muscle in phenylephrine-contracted rat aorta via the up-regulation of cGMP level to show its vasorelaxation activity [62]. Finally, among in vitro screening of antiviral drugs for treating hand, foot, and mouth disease (HFMD) infection, CF aqueous extract (at 0.4 μg mL $^{-1}$) for 2 days significantly inhibited CVA16 replication in cellular level [63].

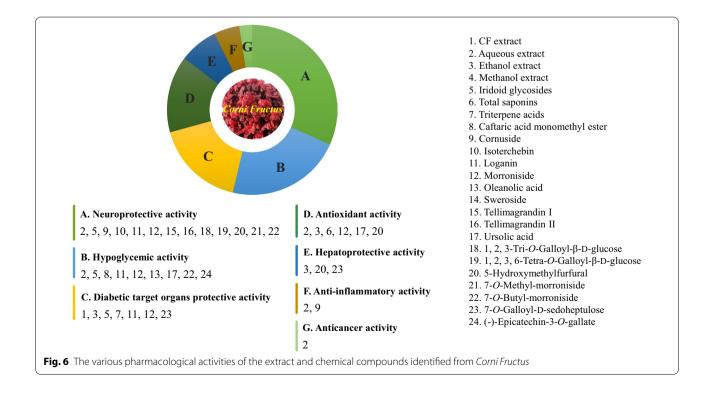
Conclusion

CF is recognized as a fundamental constituent part of tonifying Yin and Yang prescription because of its harmonious and complementary features according to the basic theory of TCM. It possesses the properties of sour and astringent. Firstly, sour and sweet herbs can be combined to nourish Yin, it can act as the sovereign and ministerial drug among *Radix Rehmanniae Praeparata*, *Dioscoreae Rhizoma*, *Lycii Fructus*, *Ligustri Lucidi Fructus*, *Schisandrae Chinensis Fructus*. Also, sour and astringent properties exhibit their function of astringing and storing. It also behaves as the sovereign and the ministerial drug that combines with *Euryales Semen*, *Sepiae Endoconcha*, *Mantidis Oötheca*, *Rubi Fructus*, *Paeoniae Radix Alba* to

treat spermatorrhea, urorrhagia, metrorrhagia and metrostaxis, and excessive perspiration. Finally, CF can be as the adjuvant and conductant drug to alleviate warm and dry features of Yang-reinforcing drugs.

Chemical constituents from terpenoids, flavonoids, tannins, and furans exhibited diverse biological activities, including hypoglycemic, neuroprotective, heart-protective, hepatoprotective, nephroprotective, testis-protective activities. Pharmacological activities are outlined in Fig. 6. In these studies, bioactive components from CF mainly alleviated the damage of target organs by antioxidant activity, anti-inflammatory activity, and anti-apoptosis activity, i.e., up-regulating the expressions and activities of antioxidant enzymes, down-regulating the levels of cytokines and chemokines, and modulating the abnormal expressions of apoptotic death associated proteins.

Hypoglycemic activity and alleviating diabetic target organs damage are critical pharmacological activities among the broad spectrum of pharmacological activities of CF. Morroniside, loganin, oleanolic acid, ursolic acid, and 7-O-galloyl-D-sedoheptulose exhibited the similar efficacy compared with the conventional oral hypoglycemic drugs (acarbose and metformin). In vivo studies, they reduced serum glucose levels and alleviated unusual symptoms caused by diabetes. In cellular assays, they protected pancreatic β cell from oxidative damage, increased insulin release, improved insulin resistance, displayed α -glucosidase inhibition activity, and



suppressed liver gluconeogenesis. Also, compounds alleviated the high-glucose triggered target organs damage by attenuating oxidative stress, inflammation, and apoptosis, finally kept the essential function of target organs stable. CF has also been widely used to treat DM in clinical work. For example, Jingui Shenqi Wan and Liuwei Dihuang Wan are two classic Chinese medicinal formulae which contain CF. Clinical trials indicated that Jingui Shenqi Wan and Liuwei Dihuang Wan could decrease serum glucose levels, alleviate typical DM symptoms and repair target organs injury [64–67]. Diverse anti-diabetes and anti-diabetic complication pharmacological activities make CF a potential herb to become the complementary drug for treating DM.

Another significant biological activity is the neuroprotection. In cerebral ischemia rat model and neurodegenerative disorder cellular model, iridoid glycosides (e.g., morroniside) and 5-hydroxymethylfurfural increased the number of new mature neurons and blood vessels and exerted anti-oxidative stress, anti-inflammation, and anti-apoptosis properties. In cerebral ischemia rat model and multiple sclerosis rats and mice models, iridoid glycosides also enhanced the levels of brain-derived neurotrophic factor and nerve growth factor. Current studies showed that the pathogenic mechanisms of neurodegenerative diseases have the close relationship with autophagy deficiency and abnormal proteins aggregate clearance dysfunction [68, 69]. In addition to the antiapoptotic activity, pharmacological activities of CF on the regulation of autophagy can be further explored. Furthermore, many classic Chinese medicinal formulae have been used to treat neurological disorders belonging to liver and kidney deficiency [70–72]. For example, Liuwei Dihuang Wan treats insomnia, Zuogui Wan (左 归丸) treats epilepsy and vertigo, Dabu Yinjian (大补阴 煎) treats a headache, Zuogui Wan and Dihuang Yinzi (地 黄饮子) treats stroke, and Huanshao Dan (还少丹) treats dementia. CF plays a vital role in nourishing liver and kidney Yin in these Chinese medicinal formulae.

However, about 90 compounds have been isolated and identified from CF, only 18% compounds are further assayed for their pharmacological activities in vivo and in vitro. It indicates that pharmacological activities of the remaining 90% chemical components are still unknown yet. Moreover, current studies do not provide enough evidence to verify the drug binding sites of active ingredients of CF. For example, it is difficult to judge whether these active ingredients bind the G protein coupled receptor, ion channels, transmembrane receptor kinases, or nuclear receptors to work. Therefore, more systematic and detailed pharmacological studies on CF need to be fulfilled in the future.

Abbreviations

ACh: acetyl choline; AGEs: advances glycation endproducts; ALP: alkaline phosphatase; ALT: alanine aminotransferase; AST: aspartate aminotransferase; ATF4: activating transcription factor 4; BACE1: b-site amyloid precursor protein cleaving enzyme 1; Bax: bcl-2-associated X; Bcl-2: B-cell lymphoma-2; BDNF: brain-derived neurotrophic factor; BMDM: bone marrow-derived macrophages; BrdU: bromodeoxyuridine; CAT: catalase; CEL: Ne-(carboxyethyl) lysine; ChE: cholinesterase; CHOP: C/EBP homologous protein; CMA: Ne-(carboxymethyl)arginine; CML: Ne-(carboxymethyl)lysine; Col V: collagen V; COX-2: cyclooxygenase-2; CTGF: connective tissue growth factor; CVA16: Coxsackie virus A group 16 strain; Cyt c: cytochrome C; DCs: dendritic cells; E2: 17β-estradiol; E3: estrone; ECE: endothelin converting enzyme; ECM: extracellular matrix; ER+: estrogen receptor-positive; ERK1/2: extracellularsignal-related kinase 1/2; ET-1: endothelin-1; FN: fibronectin; GAP-43: growthassociated protein-43; GC-MS: gas chromatography-mass spectrometry; GPX: glutathione peroxidase; GSH: glutathione; GSP: glycated serum protein; GSSG: glutathione disulfide; HK-2: human renal proximal tubular epithelial cells; HO-1: heme oxygenase-1; HUVECs: human umbilical vein endothelial cells; ICAM-1: intercellular adhesion molecule-1; iNOS: inducible nitric oxide synthase; IFN: interferon; IL: interleukin; JNK: c-Jun N-terminal kinase; LPS: lipopolysaccharide; MAPK: mitogen-activated protein kinase; MCAO: middle cerebral artery occlusion; MCP-1: monocyte chemoattractant protein 1; MDA: malondialdehyde; MITF-M: microphthalmia-associated transcription factor-M; MMP: mitochondrial membrane potential; NF-kB: nuclear factorkappa B; NFATc1: nuclear factor of activated T cells cytoplasmic 1; NGF: nerve growth factor; NO: nitric oxide; OSCAR: osteoclast-associated receptor; OVA: ovalbumin: p-eIF2q: p-eukarvotic initiation factor 2 alpha: PAFCs: pulmonarv artery endothelial cells; PEPCK: phosphoenolpyruvate carboxykinase; PERK: protein kinase R (PKR)-like endoplasmic reticulum kinase; PGE₂: prostaglandin E2; PLB: phospholamban; p.o.: per os; RAGE: receptor of AGE; RANKL: receptor activator of nuclear factor kappa-B ligand; ROS: reactive oxygen species; SERCA2a: sarcoplasmic reticulum Ca²⁺-ATPase 2a; SOD: superoxide dismutase; SPHK1: sphingosine kinase 1; SREBP-1&2: sterol regulatory element binding protein-1&2; STIM1: sensor protein stromal interaction molecule 1; STZ: streptozotocin; SYP: synaptophysin; TBARS: thiobarbituric acid-reactive substance; TC: triglyceride; TG: total cholesterol; TGF: transforming growth factor; TNF-a: tumor necrosis factor-α; TRAP: tartrate-resistant acid phosphatase; Trk A: tyrosine receptor kinase A; TRP-1&2: tyrosinase-related protein-1&2; VCAM-1: vascular cell adhesion molecule-1; VEGF: vascular endothelial growth factor.

Authors' contributions

YD conducted literature searches, extracted and analyzed data and drafted the manuscript and prepared tables and figures. ZLF contributed to the proofreading of chemical constituents and structures. FSW contributed to the draft of the fundamental theories of traditional Chinese medicine of the review. HBC contributed to the revisions of the manuscript. JHL designed the study, developed and revised the manuscript and is the corresponding author. All authors read and approved the final manuscript.

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The authors declare that they have no competing interests.

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References

- Chinese Pharmacopoeia Commission. People's Republic of China pharmacopoeia 2015 edition, 1st part. Beijing: China Medical Science Press; 2015. p. 27–8 (in Chinese).
- 2. Xiang CK. Research progress on resources and active components of *Cornus officinalis*. Hebei Med J. 2016;38:1886–9 (in Chinese).
- Song XY, Chen Q, Qi XY. Effect of Liuwei Dihuang pill on erythrocyte aldose reductase activity in early diabetic nephropathy patients. Chin J Integr Tradit West Med. 2004;24:1087–90.
- Liu XD, Fu J, Feng MZ, Zhang ZH. Effect of Jinggui Shenqi pill combined with nifedipine for the treatment of elderly hypertensive patients with spleen–kidney Yang deficiency syndrome. China J Chin Mater Med. 2015;40:4908–12 (in Chinese).
- Wang Y, Wang C. Three cases of clinical application of Guchong Decoction. China J Tradit Chin Med Pharm. 2010;25:2041–3 (in Chinese).
- Hatano T, Ogawa N, Kira R, Yasuhara T, Okuda T. Tannins of cornaceous plants. I. Cornusiins A, B and C, dimeric monomeric and trimeric hydrolyzable tannins from *Cornus officinalis*, and orientation of valoneoyl group in related tannins. Chem Pharm Bull. 1989;37:2083–90.
- Hatano T, Ogawa N, Kira R, Yasuhara T, Okuda T. Tannins of cornaceous plants. II. Cornusiins D, E and F, new dimeric and trimeric hydrolyzable Tannins from Cornus officinalis. Chem Pharm Bull. 1989;37:2665–9.
- 8. Wu Y, Wang X, Shen B, Kang L, Fan E. Extraction, structure, and bioactivities of the polysaccharides from *Fructus corni*. Recent Pat Food Nutr Agric. 2013;5:57–61.
- Yin X, You Q, Jiang Z, Zhou X. Optimization for ultrasonic-microwave synergistic extraction of polysaccharides from *Cornus officinalis* and characterization of polysaccharides. Int J Biol Macromol. 2016;83:226–32.
- Chen H, Zhang HJ, Zhang MJ, Xiao ZH. Orthogonal test for optimization of extraction condition for volatile components of *Cornus officinalis* and extract analysis by GC–MS. J Third Mil Med Univ. 2007;29:1079–82 (in Chinese).
- Li GY, Yao YX, Ding X. Studies on chemistry component and the biological activity of petroleum ether extraction from pre- and post-processed of Cornus officinalis. Zhong Yao Cai. 2010;33:192–5 (in Chinese).
- Wen YY, Ren Q, Zhang GM, Li AN, Ding ZE. Analysis of the essential oils from Cornus officinalis by simultaneous distillation extraction coupled with gas chromatography–mass spectrometry. Food Ferment Ind. 2010;36:166–70
- Han Y, Jung HW, Park YK. Selective therapeutic effect of Cornus officinalis fruits on the damage of different organs in STZ-induced diabetic rats. Am J Chin Med. 2014;42:1169–82.
- He K, Song S, Zou Z, Feng M, Wang D, Wang Y, Li X, Ye X. The hypoglycemic and synergistic effect of loganin, morroniside, and ursolic acid isolated from the fruits of *Cornus officinalis*. Phytother Res. 2015;30:283–91.
- Lin MH, Liu HK, Huang WJ, Huang CC, Wu TH, Hsu FL. Evaluation of the potential hypoglycemic and beta-cell protective constituents isolated from *Corni Fructus* to tackle insulin-dependent diabetes mellitus. J Agric Food Chem. 2011;59:7743–51.
- He LH. Comparative study on alpha-glucosidase inhibitory effects of total iridoid glycosides in the crude products and the wine-processed products from Cornus officinalis. Yakugaku Zasshi. 2011;131:1801–5.

- Park CH, Noh JS, Tanaka T, Uebaba K, Cho EJ, Yokozawa T. The effects of Corni Fructus extract and its fractions against α-glucosidase inhibitory activities in vitro and sucrose tolerance in normal rats. Am J Chin Med. 2011;39:367–80.
- Chen CC, Hsu CY, Chen CY, Liu HK. Fructus Corni suppresses hepatic gluconeogenesis related gene transcription, enhances glucose responsiveness of pancreatic beta-cells, and prevents toxin induced beta-cell death. J Ethnopharmacol. 2008;117:483–90.
- 19. Yokozawa T, Kang KS, Park CH, Noh JS, Yamabe N, Shibahara N, Tanaka T. Bioactive constituents of *Corni Fructus*: the therapeutic use of morroniside, loganin, and 7-O-galloyl-o-sedoheptulose as renoprotective agents in type 2 diabetes. Drug Discov Ther. 2010;4:223–34.
- Yamabe N, Kang KS, Goto E, Tanaka T, Yokozawa T. Beneficial effect of Corni Fructus, a constituent of Hachimi-jio-gan, on advanced glycation end-product-mediated renal injury in streptozotocin-treated diabetic rats. Biol Pharm Bull. 2007;30:520–6.
- Yamabe N, Kang KS, Park CH, Tanaka T, Yokozawa T. 7-O-galloyl-p-sedoheptulose is a novel therapeutic agent against oxidative stress and advanced glycation endproducts in the diabetic kidney. Biol Pharm Bull. 2009;32:657-64.
- Lv X, Dai G, Lv G, Chen Y, Wu Y, Shen H, Xu H. Synergistic interaction of effective parts in *Rehmanniae Radix* and *Cornus officinalis* ameliorates renal injury in C57BL/KsJ-db/db diabetic mice: involvement of suppression of AGEs/RAGE/SphK1 signaling pathway. J Ethnopharmacol. 2016;185:110–9.
- 23. Jiang WL, Zhang SP, Hou J, Zhu HB. Effect of loganin on experimental diabetic nephropathy. Phytomedicine. 2012;19:217–22.
- Ma W, Wang KJ, Cheng CS, Yan GQ, Lu WL, Ge JF, Cheng YX, Li N. Bioactive compounds from *Cornus officinalis* fruits and their effects on diabetic nephropathy. J Ethnopharmacol. 2014;153:840–5.
- Xu HQ, Hao HP. Effects of iridoid total glycoside from Cornus officinalis on prevention of glomerular overexpression of transforming growth factor beta 1 and matrixes in an experimental diabetes model. Biol Pharm Bull. 2004;27:1014–8.
- Qi MY, Liu HR, Dai DZ, Li N, Dai Y. Total triterpene acids, active ingredients from *Fructus Corni*, attenuate diabetic cardiomyopathy by normalizing ET pathway and expression of FKBP12.6 and SERCA2a in streptozotocin-rats. J Pharm Pharmacol. 2008;60:1687–94.
- Chen Y, Wu Y, Gan X, Liu K, Lv X, Shen H, Dai G, Xu H. Iridoid glycoside from *Cornus officinalis* ameliorated diabetes mellitus-induced testicular damage in male rats: involvement of suppression of the AGEs/RAGE/p38 MAPK signaling pathway. J Ethnopharmacol. 2016;194:850–60.
- 28. Pi WX, Feng XP, Ye LH, Cai BC. Combination of morroniside and diosgenin prevents high glucose-induced cardiomyocytes apoptosis. Molecules.
- 29. Wang W, Sun F, An Y, Ai H, Zhang L, Huang W, Li L. Morroniside protects human neuroblastoma SH-SY5Y cells against hydrogen peroxide-induced cytotoxicity. Eur J Pharmacol. 2009;613:19–23.
- Liu Y, Liu H, Zhang J. Total saponins of Cornus officinalis Sieb. ameliorates the endothelium dependent relaxation of mesenteric artery by regulating nitric oxide release in streptozotocin-induced diabetic rats. Zhong Nan Da Xue Xue Bao Yi Xue Ban. 2012;37:757–64.
- 31. Peng Q, Wei Z, Lau BH. *Fructus corni* enhances endothelial cell antioxidant defenses. Gen Pharmacol. 1998;31:221–5.
- 32. Yu HH, Hur JM, Seo SJ, Moon HD, Kim HJ, Park RK, You YO. Protective effect of ursolic acid from *Cornus officinalis* on the hydrogen peroxide-induced damage of HEI-OC1 auditory cells. Am J Chin Med. 2009;37:735–46.
- Hwang KA, Hwang YJ, Song J. Antioxidant activities and oxidative stress inhibitory effects of ethanol extracts from *Cornus officinalis* on raw 264.7 cells. BMC Complement Altern Med. 2016;16:196.
- Cao G, Cai H, Cai B, Tu S. Effect of 5-hydroxymethylfurfural derived from processed *Cornus officinalis* on the prevention of high glucose-induced oxidative stress in human umbilical vein endothelial cells and its mechanism. Food Chem. 2013;140:273–9.
- Xu HQ, Hao HP, Zhang X, Pan Y. Morroniside protects cultured human umbilical vein endothelial cells from damage by high ambient glucose. Acta Pharmacol Sin. 2004;25:412–5.
- Kang DG, Moon MK, Lee AS, Kwon TO, Kim JS, Lee HS. Cornuside suppresses cytokine-induced proinflammatory and adhesion molecules in the human umbilical vein endothelial cells. Biol Pharm Bull. 2007;30:1796–9.

- Chu Q, Hashimoto K, Satoh K, Wang Q, Sakagami H. Effect of three herbal extracts on NO and PGE2 production by activated mouse macrophagelike cells. In Vivo. 2009;544:537–44.
- Choi YH, Jin GY, Li GZ, Yan GH. Cornuside suppresses lipopolysaccharideinduced inflammatory mediators by inhibiting nuclear factor-kappa B activation in RAW 264.7 macrophages. Biol Pharm Bull. 2011;34:959–66.
- Chu Q, Satoh K, Kanamoto T, Terakubo S, Nakashima H, Wang Q, Sakagami H. Antitumor potential of three herbal extracts against human oral squamous cell lines. Anticancer Res. 2009;29:3211–9.
- Telang NT, Li G, Sepkovic DW, Bradlow HL, Wong GY. Anti-proliferative effects of Chinese herb Cornus officinalis in a cell culture model for estrogen receptor-positive clinical breast cancer. Mol Med Rep. 2012;5:22–8.
- Telang N, Li G, Katdare M, Sepkovic D, Bradlow L, Wong G. Inhibitory effects of Chinese nutritional herbs in isogenic breast carcinoma cells with modulated estrogen receptor function. Oncol Lett. 2016;12:3949–57.
- 42. Chang JS, Chiang LC, Hsu FF, Lin CC. Chemoprevention against hepatocellular carcinoma of *Cornus officinalis* in vitro. Am J Chin Med. 2004;32:717–25.
- Jeong EJ, Kim TB, Yang H, Kang SY, Kim SY, Sung SH, Kim YC. Neuroprotective iridoid glycosides from *Cornus officinalis* fruits against glutamate-induced toxicity in HT22 hippocampal cells. Phytomedicine. 2012;19:317–21.
- Wang X, Liu J, Jin N, Xu D, Wang J, Han Y, Yin N. Fructus corni extractinduced neuritogenesis in PC12 cells is associated with the suppression of stromal interaction molecule 1 expression and inhibition of Ca²⁺ influx. Exp Ther Med. 2015;9:1773–9.
- Bhakta HK, Park CH, Yokozawa T, Tanaka T, Jung HA, Choi JS. Potential anticholinesterase and β-site amyloid precursor protein cleaving enzyme 1 inhibitory activities of cornuside and gallotannins from *Cornus officinalis* fruits. Arch Pharm Res. 2017;40:836–53.
- Yao RQ, Zhang L, Wang W, Li L. Cornel iridoid glycoside promotes neurogenesis and angiogenesis and improves neurological function after focal cerebral ischemia in rats. Brain Res Bull. 2009;79:69–76.
- Wang W, Xu J, Li L, Wang P, Ji X, Ai H, Zhang L, Li L. Neuroprotective effect of morroniside on focal cerebral ischemia in rats. Brain Res Bull. 2010;83:196–201.
- 48. Yin L, Chen Y, Qu Z, Zhang L, Wang Q, Zhang Q, Li L. Involvement of JAK/ STAT signaling in the effect of cornel iridoid glycoside on experimental autoimmune encephalomyelitis amelioration in rats. J Neuroimmunol. 2014;274:28–37
- 49. Zhao L, Ding Y, Zhang L, Li L. Cornel iridoid glycoside improves memory ability and promotes neuronal survival in fimbria–fornix transected rats. Eur J Pharmacol. 2010;647:68–74.
- Qu Z, Zheng N, Zhang Y, Zhang L, Liu J, Wang Q, Yin L. Preventing the BDNF and NGF loss involved in the effects of cornel iridoid glycoside on attenuation of experimental autoimmune encephalomyelitis in mice. Neurol Res. 2016;38:831–7.
- Ding X, Wang MY, Yao YX, Li GY, Cai BC. Protective effect of 5-hydroxymethylfurfural derived from processed *Fructus Corni* on human hepatocyte L02 injured by hydrogen peroxide and its mechanism. J Ethnopharmacol. 2010;128:373–6.
- Wang MY, Zhao FM, Peng HY, Lou CH, Li Y, Ding X, Yu XY, Yang GM, Xu DQ, Jiang LH, Zhang X, Ye LH, Cai BC. Investigation on the morphological protective effect of 5-hydroxymethylfurfural extracted from wineprocessed *Fructus corni* on human L02 hepatocytes. J Ethnopharmacol. 2010;130:424–8.
- Jiang ZQ, Ma YX, Li MH, Zhan XQ, Zhang X, Wang MY. 5-Hydroxymethylfurfural protects against ER stress-induced apoptosis in GalN/TNF-αinjured L02 hepatocytes through regulating the PERK-elF2α signaling pathway. Chin J Nat Med. 2015;13:896–905.
- Park CH, Noh JS, Tanaka T, Roh SS, Lee JC, Yokozawa T. Polyphenol isolated from Corni Fructus, 7-O-galloyl-p-sedoheptulose, modulates advanced glycation endproduct-related pathway in type 2 diabetic db/db mice. Arch Pharmacal Res. 2015;38:1270–80.
- Lee NH, Seo CS, Lee HY, Jung DY, Lee JK, Lee JA, Song KY, Shin HK, Lee MY, Seo YB, Kim H, Ha H. Hepatoprotective and antioxidative activities of Cornus officinalis against acetaminophen-induced hepatotoxicity in mice. Evid Based Complement Altern Med. 2012;2012:1–8.
- Sun H, Li L, Zhang A, Zhang N, Lv H, Sun W, Wang X. Protective effects of sweroside on human MG-63 cells and rat osteoblasts. Fitoterapia. 2013;84:174–9.

- 57. Kim JY, Kim YK, Choi MK, Oh J, Kwak HB, Kim JJ. Effect of *Cornus Officinalis* on receptor activator of nuclear factor-kappaB ligand (RANKL)-induced osteoclast differentiation. J Bone Metab. 2012;19:121.
- 58. An YA, Hwang JY, Lee JS, Kim YC. *Cornus officinalis* methanol extract upregulates melanogenesis in melan-a cells. Toxicol Res. 2015;31:165–72.
- Liu X, Zeng YQ, Liang YZ, Zou C, Liu H, Qiu F, Liang CL, Jin XW, Su ZR, Dai Z. Medicinal herbs *Fructus corni* and *Semen cuscutae* suppress allograft rejection via distinct immune mechanisms. Oncotarget. 2016;7:35680.
- Cho K, Lee HJ, Lee SY, Woo H, Lee MN, Seok JH, et al. Oleanolic acid and ursolic acid derived from *Cornus officinalis* Sieb. et Zucc. suppress epidermal growth factor- and phorbol ester-induced MUC5AC mucin production and gene expression from human airway epithelial cells. Phytother Res. 2011;25:760–4.
- Kim SH, Kim BK, Lee YC. Effects of *Corni fructus* on ovalbumin-induced airway inflammation and airway hyper-responsiveness in a mouse model of allergic asthma. J Inflamm. 2012;9:9.
- 62. Kang DG, Choi DH, Lee JK, Lee YJ, Moon MK, Yang SN, Kwon TO, Kwon JW, Kim JS, Lee HS. Endothelial NO/cGMP-dependent vascular relaxation of cornuside isolated from the fruit of *Cornus officinalis*. Planta Med. 2007;73:1436–40.
- Song JH, Park K, Shim A, Kwon BE, Ahn JH, Choi YJ, Kim JK, Yeo SG, Yoon K, Ko HJ. Complete sequence analysis and antiviral screening of medicinal plants for human Coxsackievirus A16 isolated in Korea. Osong Public Health Res Perspect. 2015;6:52–8.
- 64. Wu HZ. The clinical effect of Jinkui Shenqi pill on type 2 diabetes. Pharmacol Clin Chin Mater Med. 2013;29:191–3 (in Chinese).
- Liu ZW. Curative effect valuation on Jingui Senqi pill in the treatment of diabetic nephropathy. Chin J Basic Med Tradit Chin Med. 2014;20(821–2):31.
- Li X, Li Y. Clinical study on Liuwei Dihuang pills combined with liraglutide and metformin in treatment of type 2 diabetes mellitus. Drugs Clin. 2016;31:1146–50 (in Chinese).
- He K, Zhu LH, Lu XW. Clinical observation of Liuwei Dihuang pills combined with metformin in the treatment of type 2 diabetes. Chin Tradit Pat Med. 2016;38:50–2.
- 68. Menzies FM, Fleming A, Rubinsztein DC. Compromised autophagy and neurodegenerative diseases. Nat Rev Neurosci. 2015;16:345–57.
- 69. Nah J, Yuan J, Jung YK. Autophagy in neurodegenerative diseases: from mechanism to therapeutic approach. Mol Cells. 2015;38:381–9.
- 70. Zhou ZY. Chinese internal medicine. Beijing: China Press of Traditional Chinese Medicine; 2003. p. 157, 176, 183, 308, 316, 327 (in Chinese).
- Lin SK, Yan SH, Lai JN, Tsai TH. Patterns of Chinese medicine use in prescriptions for treating Alzheimer's disease in Taiwan. Chin Med. 2016;11:12.
- Yeung WF, Chung KF, Zhang NL, Zhang SP, Yung KP, Chen PX, Ho YY. Identification of Chinese medicine syndromes in persistent insomnia associated with major depressive disorder: a latent tree analysis. Chin Med. 2016;11:4.
- 73. Cai H, Cao G, Cai B. Rapid simultaneous identification and determination of the multiple compounds in crude *Fructus Corni* and its processed products by HPLC–MS/MS with multiple reaction monitoring mode. Pharm Biol. 2013;51:273–8.
- 74. Zhang YE, Liu EH, Li HJ, Li P. Chemical constituents from the fruit of *Cornus officinalis*. Chin J Nat Med. 2009;7:365–7 (in Chinese).
- Han SY, Pan Y, Ding G, Cai BC. Application of 1H-NMR and 13C-NMR spectra in the structure identification of iridoids from *Cornus officinalis*. Chin Arch Tradit Chin Med. 2004;22:56–9 (in Chinese).
- Park JY, Han AR, Kil YS, Kang U, Kim SH, Nam SJ, Seo EK. A new secoiridoid glycoside from the fruits of *Cornus officinalis* (Cornaceae). Nat Prod Res. 2016;30:1504–10.
- 77. He J, Ye XS, Wang XX, Yang YN, Zhang PC, Ma BZ, Zhang WK, Xu JK. Four new iridoid glucosides containing the furan ring from the fruit of *Cornus officinalis*. Fitoterapia. 2017;120:136–41.
- Han SY, Pan Y, Yang GM, Cai BC. Research on components of *Cornus offici-nalis* extracted by supercritical carbon dioxide. Zhongguo Zhong Yao Za Zhi. 2003;28:1148–50, 83 (in Chinese).
- Kim DK, Kwak JY. A furan derivative from Comus officinalis. Arch Pharm Res. 1998;21:787–9.
- 80. Xie X, Wang R, Shi Y. Chemical constituents from the fruits of *Cornus officinalis*. Biochem Syst Ecol. 2012;45:120–3.

- 81. Yang L, Wang Z, Huang L. Isolation and structural characterization of a polysaccharide FCAP1 from the fruit of *Cornus officinalis*. Carbohydr Res. 2010;345:1909–13.
- 82. Wang LL, Zhang T, Chen SQ, Shang CL. Study on the correlation between the quality of *Cornus officinalis* and the contents of the inorganic elements in the planting soil. Zhong Yao Cai. 2011;34:1167–72 (in Chinese).
- 83. Hsu JH, Wu YC, Liu IM, Cheng JT. Release of acetylcholine to raise insulin secretion in Wistar rats by oleanolic acid, one of the active principles contained in *Cornus officinalis*. Neurosci Lett. 2006;404:112–6.
- 84. Gu H, Jiang Z, Wang M, Jiang H, Zhao F, Ding X, Cai B, Zhan Z. 5-Hydroxy-methylfurfural from wine-processed *Fructus corni* inhibits hippocampal neuron apoptosis. Neural Regen Res. 2013;8:2605–14.
- Zhang QC, Zhao Y, Bian HM. Antiplatelet activity of a novel formula composed of malic acid, succinic acid and citric acid from *Cornus officinalis* fruit. Phytother Res. 2013;27:1894–6.

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